



THE OHIO STATE
UNIVERSITY

Animal Care and Use Newsletter

In This Issue:

- New IACUC Chair
- New Policies
- Revised Policies
- Reminders
- Coming Soon...
- E-Protocol Tips

August
2013

IACUC
Institutional Animal Care and Use Committee
ULAR
University Laboratory Animal Resources
ORRP
Office of Responsible Research Practices

New IACUC Chair

Dr. Janet Weisenberger, the Senior Associate Vice President for Research, announced at the June IACUC meeting that Dr. Donna McCarthy would be stepping down from her term as the IACUC Chair. Dr. McCarthy came to OSU in January of 2007 as the Associate Dean for Research and Mildred E. Newton Char in the College of Nursing. She joined the IACUC in the summer of 2007 and assumed the role of Chair in July of 2009.

Dr. McCarthy's research program at OSU (funded by an R15 and R01) examined the interactions of muscle wasting, heart dysfunction, and depressed mood in animal models of cancer-related fatigue. During her tenure here, McCarthy has been commuting to Madison, WI where her family still lives. She has accepted a position at Marquette University in Milwaukee, a mere 90 miles from her home, to shorten her commute time. We wish Dr. McCarthy the best of luck and thank her for her service, dedication and leadership.

Dr. Peter J. Reiser is the new Chair effective July 1, 2013. Dr. Reiser holds dual appointments as a professor of oral biology in the College of Dentistry and a professor of physiology and cell biology in the College of Medicine. Dr. Reiser's research includes projects that examine craniofacial muscles in a broad range of vertebrate species, focusing on how contractile protein isoforms regulate contractile properties of striated muscle. He also studies physiological changes in muscle associated with cancer cachexia.

Dr. Reiser received his PhD in Physiology at OSU in 1981 and returned as a faculty member in 1992. He has been a member of the IACUC since 1995. We look forward to working with Dr. Reiser as the new IACUC Chair.



Resources

Animal Care & Use
Newsletter Online:
<http://orrr.osu.edu/iacuc>

e-Protocol:
<https://eprotocol.osu.edu>

University Laboratory
Animal Resources:
<http://ular.osu.edu>

Office of Responsible
Research Practices:
<http://orrr.osu.edu>

Institutional Biosafety
Committee:
<http://orrr.osu.edu/ibc>

Office of Environmental
Health and Safety:
<http://ehs.osu.edu>

New IACUC Policies

- **New - Euthanasia of Rodent Fetuses and Neonates**: This policy was previously part of the CO₂ Euthanasia of Rodents Including Guinea Pigs Policy (see revised policy below) and has been separated out into a new stand-alone policy.
- **New - Validate Sanitization Effectiveness**: Sanitization verification for **primary enclosures and husbandry equipment** (i.e., all housing items such as cages, wire feeders, enrichment devices, water bottles, data collection equipment inside the cage, etc.) must be monitored on a semi-annual basis. In most cases, if automated sanitization is provided by ULAR (i.e., cage wash, tunnel wash) then verification is monitored by ULAR and no further action is required. If housing enclosures and/or equipment are manually sanitized by the lab then sanitization methods must be verified every six months, and documentation of results must be available for review during semiannual inspections.

Sanitization verification for anything that is **not used for housing or found inside the cage** (e.g., non-permeable work surfaces, behavioral equipment, secondary enclosures, etc.) must be validated initially, then again *only after any change in the cleaning process*. Documentation of initial validation must be available for review during semiannual inspections.

Important Information about Cleaning Products: If you use any of these products and follow the manufacturer's instructions, including proper dilution and contact time, then no further validation is needed. The following products have been validated and approved:



Wex-Cide

Spor-Klenz

Opti-Cide

Policies Revised in 2013

All of the policies developed by the OSU Institutional Animal Care and Use Committee to assist investigators and staff with their research are available on the [Policies Page on the ORRP Website](#). These policies are reviewed and updated on a regular basis to ensure that they still meet regulatory requirements.

- **Revised - [Vertebrate Animal Field Studies](#)**: Formally the “Wildlife Research” Policy, this policy was revised to be more concise and move the requirements closer to the beginning of the policy. **Important Addition to the Policy: A Field Safety Plan must be in place prior to protocol approval. Contact Environmental Health and Safety for assistance with the plan.**
- **Revised - [Transfer of Animals to Holding Protocol](#)**: The revision of this policy explicitly states procedures that may or may not be performed on animals maintained on the IACUC Holding Protocol and defines the roles and responsibilities of all personnel involved in the transfer process. **Important Addition to the Policy: Animals remaining on the IACUC Holding Protocol after 60 days will be euthanized unless other arrangements have been approved by the IACUC.**
- **Revised - [Documentation of Husbandry Procedures](#)**: This policy was updated to **require**, rather than recommend, documentation of activities performed by lab personnel, such as providing study-specific food or water, changing cages on a special schedule, etc. ULAR provides “**purple cards**” that may be placed behind the cage card and be used to record these activities performed by lab personnel. Documentation may also be recorded at the room level instead of the cage level.
- **Revised - [Housing Requirements for Animals](#)**: This policy was revised to include definitions of the housing options at OSU, an improved description of the requirements for all housing locations, and a better description of exceptions to the [Guide](#) standards.
- **Revised - [CO₂ Euthanasia of Rodents Including Guinea Pigs](#)**: This policy was updated to remain consistent with the recommendations of the *2013 AVMA Guidelines on Euthanasia*. In addition, the section on Euthanasia of Rodent Fetuses and Neonates was separated out into a stand-alone policy (see above). **Important Additions to the Policy: Prefilling of the chambers is unacceptable, the flow of CO₂ must be maintained for at least ONE (1) minute after respiratory arrest, and cervical dislocation must not be performed in rats weighing more than 200 grams.**

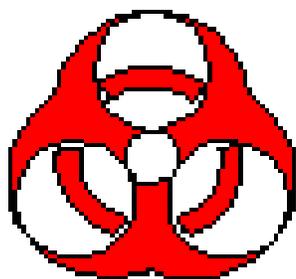


**R
E
M
I
N
D
E
R
S**

- **Confirmation of death after euthanasia** - confirmation of death is **required** after CO₂ euthanasia prior to disposal of carcasses. A secondary physical method of euthanasia **must** be performed following any euthanasia from inhaled agents. Acceptable secondary methods for confirmation of death include creation of a pneumothorax, removal of a vital organ, decapitation, exsanguination, or cervical dislocation.
- **Weaning extensions and breeding schemes** - weaning extensions at or before 28 days are only approved for **Monogamous Pairing** cages. You may only have **ONE (1)** adult male and **ONE (1)** adult female in a cage approved for a weaning extension. Otherwise, pups **must** be weaning at or before day 21 in **Trio Breeding** schemes, or pregnant females **must** be removed and placed into another cage **before** parturition in **Harem Breeding** schemes. The complete "Mouse Breeding Cages" Policy is available on the OSU Policies website [here](#).

Remember that weaning extensions must be indicated by the research staff on the green breeding cards; otherwise, ULAR staff has been instructed to treat the cage as a 21-day weaning cage.

- **Notifying ULAR when using hazardous agents** - if your lab has been assigned an OASIS (OSU Animal Safety Information Sheet) form, you are responsible for submitting notification to ULAR three business days prior to agent administration by clicking on the "Notify ULAR of Biohazard Use" link ( [Notify ULAR of Biohazard Use](#)) in e-Protocol. Assigned OASIS forms have been emailed to investigators, or can be found under the "Attachments" tab in e-Protocol as a "Supporting Document." FAQs about OASIS forms and the procedure for notifying ULAR can be found [here](#) and [here](#), respectively.



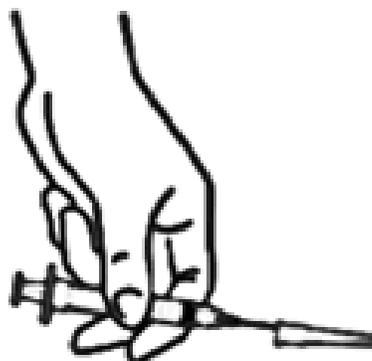
- **Using pharmaceutical-grade compounds when available** - pharmaceutical-grade compounds should be used in **all** experimental animals whenever possible. The use of non-pharmaceutical compounds in animals is acceptable **only after IACUC review and approval**. Justification for use of non-pharmaceutical-grade compounds should be based on scientific necessity and non-availability of an acceptable pharmaceutical-grade compound. Cost savings (e.g., in the case of pharmaceutical-grade **sodium pentobarbital**) is not a justification for using non-pharmaceutical-grade compounds. The OSU Policy, found [here](#), provides additional information about institution exemptions for Tribromoethanol (Avertin) and Saturated Potassium Chloride (KCl).

**R
E
M
I
N
D
E
R
S**

- **Using aseptic technique for rodent surgery** - preparation for rodent surgery includes three steps: preparation of the surgical equipment, preparation of the surgeon, and preparation of the animal. Visit [ULAR Training](#) for information about classes on aseptic techniques for rodent survival surgery.
 - **Equipment** - see Table 2 [here](#) for sterilization recommendations such as steam, dry heat or liquid chemical sterilization.
 - **Surgeon** - the IACUC [Rodent Surgery Policy](#) states the surgeon **must** wear a surgical mask, a clean surgical gown, scrub top or lab coat, and sterile or properly disinfected standard gloves. The approved method for properly disinfecting standard gloves states the gloves “must be disinfected using Sporklenz prior to starting surgery, after touching a non-sterile surface, and in-between surgeries. Allow 3-5 minutes for gloves to dry before use or wipe-down with a sterile surgical towel.”
 - **Animal** - the IACUC [Rodent Surgery Policy](#) states that hair **must** be removed and the surgical site scrubbed three times with a recommended skin disinfectant, alternating each disinfectant with a scrub of sterile water or 70% isopropyl alcohol. See Table 3 [here](#) for recommendations for skin disinfectants.
- **Avoid recapping needles** - recapping needles is dangerous because the needle could miss the cap and stab the hand holding it. You can protect yourself by planning ahead for safe handling and disposal of needles **without** recapping. If recapping is required you **must** use tongs, a recapping device or a one-hand scoop method to recap the needle (see below). EH&S has provided a Sharps Safety Fact Sheet that can be downloaded [here](#) and posted near lab sharps containers.



Recapping device



One-hand scoop

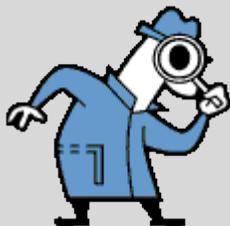
Coming Soon...

New look on the ORRP Website - watch for an updated look on the ORRP Website, which includes Animal Care and Use...

OFFICE OF RESEARCH

Office of Responsible Research Practices

[Home](#) [About Us](#) [Animal Care and Use](#) [Biosafety](#) [Human Subjects](#) [Resources](#) [Contact Us](#) [News](#)



Semiannual Inspections - mark your calendars, the next round of semiannual inspections will be starting in September.

E-Protocol Tips

Tip #1: Remember that adding new study team members is a two-step process. Any study team member can see and click the  **Propose Study Team Change** link and add new personnel, but only the investigator can see and click the  **Submit Study Team Change** link. The change in study team is **not** complete until the investigator submits the change.

Tip #2: Documentation of training is an important role of the investigator, who is ultimately responsible for all personnel listed on the protocol. One place to electronically document new training when it occurs is in the “**Experience and Training Narrative.**” Personnel can log in any time [here](#) and add information whenever a course with ULAR is completed or even when a new lab procedure is learned. It is a great place to document training and the information will not be lost.

Tip #3: Before **new** personnel can be added to a protocol they must register in e-Protocol (as well as complete all the training requirements). Registration can be completed at <http://eprotocol.osu.edu>.

Tip #4: If you have received an email assigning an OSU Animal Safety Information Sheet (OASIS) to your research, remember that an electronic copy has also been uploaded into e-Protocol. If you need to print additional copies of the OASIS and cannot find the email, the electronic file can be found under “Supporting Documents” on the “Attachments” tab next to the protocol “History” tab.

